



Review

Wild and domesticated animals as reservoirs of *Schistosomiasis mansoni* in BrazilCelina Maria Modena^a, Walter dos Santos Lima^b, Paulo Marcos Zech Coelho^{c,d,*}^a Laboratory of Health Education, René Rachou Research Center/Oswaldo Cruz Foundation, Av. Augusto de Lima 1715, Belo Horizonte, Minas Gerais, Brazil^b Department of Parasitology, Institute of Biological Science, Federal University of Minas Gerais, Av. Antonio Carlos 6627, Belo Horizonte, Minas Gerais, Brazil^c Laboratory of Schistosomiasis, René Rachou Research Center/Oswaldo Cruz Foundation, Av. Augusto de Lima 1715, Belo Horizonte, Minas Gerais, Brazil^d Hospital Santa Casa de Misericórdia, Av. Francisco Sá, Belo Horizonte, Minas Gerais, Brazil

ARTICLE INFO

Article history:

Available online 31 July 2008

Keywords:

Schistosomiasis
Reservoirs
Domesticated animals
Wild animals

ABSTRACT

Natural infection with *Schistosoma mansoni* in wild vertebrates and domesticated animals in Brazil is described in this review from an epidemiological viewpoint. Some species of wild rodents are small-sized animals, with a short expectation of life, a limited territory, and present high infection rates under natural conditions. A successful maintenance of the parasite's life cycle under artificial conditions can be achieved with *Biomphalaria glabrata*. On the other hand, despite showing low natural infection rates, cattle are very susceptible to infection under experimental conditions (using calves of Holstein lineage, cross-bred with the Gir lineage). Due to their large size (just one calf may harbor a number of worms higher than a whole colony of aquatic rodents) and their longevity, cattle are a potential reservoir for the maintenance and dissemination of the disease. There is thus a need of new studies to gain a better understanding about the actual role of these animals in the epidemiology of *S. mansoni*.

© 2008 Elsevier B.V. All rights reserved.

Schistosomiasis mansoni is a zoonosis with the human as the main and definitive host (Nelson, 1960). *Schistosomiasis japonicum*, however, is classified as an anfixenosis disease because both men and other vertebrate animals play a similar role on the maintenance of the disease in endemic areas (Nelson, 1960). Although the importance of the man for the maintenance of the life cycle of *S. mansoni* is not questioned, there are a number of studies showing that other mammals may act as source of infection (Antunes et al., 1973; Barbosa et al., 1958, 1962; Borda, 1972; Coelho et al., 1979, 1982; Fenwick, 1969; Pitchford and Visser, 1962; Piva and Barros, 1966; Martins et al., 1955; McCully and Kuger, 1969; Nelson, 1960; Schwetz, 1953). Naturally infected vertebrate animals from the orders primates, carnivora, marsupialia, artiodactyla and rodentia have already been found (D'Andrea et al., 1999, 2000, 2002; Gentile et al., 2000; Silva et al., 1992; Amorim, 1953). Sire et al. (2001) states that differences in the ecological landscape and in the epidemiological determinants are important variables to be considered in studies concerning reservoirs of schistosomiasis. Interesting observations in Guadalupe foci showed chronobiological patterns for cercarial release: early and intermediate (related mainly to human infection), and late or crepuscular (related to rodent infection). The behavior of cercariae shedding is dependent on the ecological type

of foci (Théron, 1984; Théron and Pointier, 1995). In this way, a recent paper published by Morgan et al. in 2003, in Tanzania, Africa, found *S. mansoni* female cercariae and *S. mansoni*/*Schistosomiasis rodhaini* male, hybrid males in *Biomphalaria sudanica*. This cross-breeding evidence between *S. mansoni* and *S. rodhaini* point to a possible modification of species by introgression. In Brazil, some authors debate the possibility that in some foci schistosomiasis may be maintained by humans, animals or both of them. The persistence of the focus together with the potential of exposure to the disease emphasize that new strategies of control must be considered. Too little is known regarding the role of domestic animals in the process of transmission and maintenance of the disease. As for the bovines, there are few studies that have considered either natural or experimental infection of these animals.

The first description of cattle naturally infected with *S. mansoni* was by Barbosa et al. (1962), who found four adult animals with helminthes in the mesenteries, whereas one of them showed non-viable eggs in the feces. Piva and Barros (1966) observed eggs of *S. mansoni* in the liver of three adult bovines, a phenomenon that was also reported by (Mayaudon and Power, 1970) in Venezuela. A prevalence of among 3% was detected in an epidemiologic surveillance conducted by Coelho et al. (1982) in Minas Gerais State, southeastern Brazil. The authors also showed a surprising susceptibility of calves experimentally infected with cercariae of *S. mansoni*. The authors emphasize, however, that under certain conditions, the bovine species could act permanently in an endemic focus since in one of the farms studied, four among eight bull-calves eliminated viable eggs in the faeces. Also Saede

* Corresponding author at: Laboratory of Schistosomiasis, René Rachou Research Center/Oswaldo Cruz Foundation, Av. Augusto de Lima 1715, Belo Horizonte 30190-002, Minas Gerais, Brazil. Tel.: +55 31 33497740; fax: +55 31 32953115.

E-mail address: coelhohpm@cpqrr.fiocruz.br (P.M.Z. Coelho).

et al. (1969) were able to infect calves with *S. mansoni*. Karoum and Amin (1985) evaluated the natural infection in cattle, goats, sheep, and canines in an endemic area in Sudan. From a total of 98 cattle examined through visceral inspection, only two bull-calves showed helminthes in the mesentery, including adult females of *S. mansoni* with eggs. Recently, Stothard et al. (2004) in a very intense transmission focus did not find *S. mansoni* worms in a sacrificed young heifer with a large number of *Schistosoma bovis*. Nevertheless, in spite of the absence of *S. mansoni* worms, it is important to note that the results were obtained with a single animal. It will be necessary to carry out studies in Africa with milk producing cattle (Holstein lineage and their crossbreeding descents with other lineages) that are more susceptible to *S. mansoni* infection (Modena et al., 1991). Modena et al. (1991) conducted rectal mucosal scrapes in 894 bull-calves from endemic areas of Minas Gerais, finding viable eggs in nine animals. Moreover, the authors examined 462 adult animals and none were positive to *S. mansoni* eggs in the faeces or mucosa. All positive animals belonged to small-holdings which were part of a subsistence economy. The laborers working on these farms were mostly family members. Bioecological conditions for the transmission of schistosomiasis were found in all surveyed properties. In a malacological survey of the area *Biomphalaria glabrata* specimens positive to cercariae and/or sporocysts of *S. mansoni* were observed. The human population showed the presence of *S. mansoni* eggs in the faeces. Modena et al. (1991) also showed that young animal (calves) of the Holstein lineage were very susceptible to *S. mansoni* infection and crossbreeding between Holstein and Gir lineage (most resistant) produced descendents with an intermediate degree of susceptibility.

Agricultural production at the subsistence level leads to a health profile of both human and animal population, (specifically due to zoonoses), which is inferior when compared with other forms of production. The exposure to the hazards of contracting disease is potentiated through intensive use of the land combined with a population in which there is close contact between animals and humans, and through specific forms of labor (Kloos et al., 1998). Bethony et al. (2004) previously demonstrated that schistosomiasis tends to cluster within communities, at both the neighborhood and household levels, due to the characteristic focality of risk behavior and transmission. Therefore, the effect of the presence of cattle in communities may affect the transmission of the disease. In the studies of Coelho et al. (1982) and Karoum and Amin (1985) the finding that only young animals were infected may be due to the handling conditions adopted in subsistence economy, where there is a close and frequent water use shared by human population and animals. An analysis of natural infections in non-human hosts shows that the pattern of rodent infection differs fundamentally from that of the infection in cattle. Cattle eliminate about 10% of their body weight through their faeces and thus the environmental contamination is significant. Moreover, the cattle can disseminate the parasite during breeding and daily movement.

Evaluation of the role of cattle in natural transmission should not involve drug treatment. Coelho et al. (1990) showed that the drug oxamniquine was not effective in the treatment of experimental *S. mansoni*. The authors infected experimentally 10 bull-calves with 20,000 cercariae as described by Coelho et al. (1982). Sixty-seven days after animals were treated with 15 or 30 mg/kg of oral oxamniquine and were sacrificed 30 days after treatment. There are no reports of any drug which is effective against bovine schistosomiasis in the available literature. The organization of the environment and ethological aspects of different hosts should be taken into consideration in the transmission of *S. mansoni* (Combes et al., 1994; Théron, 1984, 1989). Other domestic animals were previously evaluated as definitive hosts by means of experimental infection. Coelho et al. (1989) concluded, using experimental

infection, that *Bubalus bubalis* is a refractory host to infection with *S. mansoni*. Ocampo et al. (1981) also showed that swine (*Sus scrofa*) are naturally resistant to experimental infection. The transmission of *S. mansoni* under experimental conditions using the Bovine – *B. glabrata* – Bovine model, was carried out by Modena et al. (1993). The completion of the cycle gives evidence for the possibility of bovine species acting as a potential player in the maintenance and dissemination of the disease, independently from human population. In the same way, using wild rodents infected with *S. mansoni* and *B. glabrata* under natural conditions, it was also possible to complete the life cycle of parasite using *Nectomys squamipes* and *Holochilus brasiliensis* (Antunes et al., 1973 and Millward de Andrade et al., 1976, respectively). However, the wild rodents, because of their small size, limited territory and low life expectancy (maximum of 2 years) present limitations as an efficient reservoir for the maintenance of disease in endemic areas. In addition in all areas in which wild rodents were found with infections by *S. mansoni* it was possible to identify a human source of faecal pollution.

Considering the agricultural expansion into new areas in Brazil, there is an urgent need to make detailed studies on role of the bovine population on the maintenance of the disease in that country. It is speculated that new forms of organization of the agrarian space, with intense transit and greater confinement, may introduce cattle as a factor in the transmission of schistosomiasis in the endemic area. This pattern of cattle maintenance tends to amplify and, may be after a time, create sufficient conditions to produce new definitive hosts. However, the statement of Koskella and Lively (2007) should be pondered upon, i.e. “understanding host–parasite coevolution requires multigenerational studies in which changes in both parasite infectivity and host susceptibility are monitored”. By questioning general laws in parasitological ecology, Poulin (2007) states that in some patterns these laws are contingent, truthful only under particular circumstances, theoretical and empirical reflections previously distinguished by Mahmoud (2004), Kristt et al. (2000), Osnas and Lively (2004), Keeling et al. (2000), and Ovaskainen and Cornell (2006).

In conclusion, considering the degree of expansion of schistosomiasis in rural areas, there is an urgent need to increase and widen the studies of the role of bovines in the epidemiologic model of this disease.

References

- Amorim, J.P., 1953. Infestação experimental e natural de murídeos pelo *Schistosoma mansoni* (Nota prévia). Rev. Bras. Malariol. 5, 219–222.
- Antunes, C.M.F., Millward, A.R., Katz, N., Coelho, P.M.Z., Pellegrino, J., 1973. Role of *Nectomys squamipes* in the epidemiology of *Schistosoma mansoni* infection. Exp. Parasitol. 34, 181–188.
- Barbosa, F.S., Coelho, M.V., Abath, E.C., 1958. Infestação natural e experimental de alguns mamíferos de Pernambuco por *Schistosoma mansoni*. Rev. Bras. Malariol. Doenças Trop. 10, 137–144.
- Barbosa, F.S., Barbosa, I., Arruda, F., 1962. *Schistosoma mansoni*: natural infection of cattle in Brazil. Science 138, 831.
- Bethony, J., Williams, J.T., Brooker, S., Gazzinelli, M.F., LoVerde, P.T., Correa-Oliveira, R., Kloos, H., 2004. Exposure to *Schistosoma mansoni* infection in a rural area in Brazil. Part III. Household aggregation of water-contact behavior. Trop. Med. Int. Health. 9, 381–389.
- Borda, C.E., 1972. Infecção natural e experimental de alguns roedores pelo *Schistosoma mansoni* (Sambon, 1907). Belo Horizonte, Instituto de Ciências Biológicas da UFMG. Dissertação (Mestrado), p. 43.
- Coelho, P.M.Z., Dias, M., Mayrink, W., et al., 1979. Wild reservoirs of *Schistosoma mansoni* from Caratinga, an endemic area of Minas Gerais State. Brazil. Am. J. Trop. Med. Hyg. 28, 163–164.
- Coelho, P.M.Z., Nogueira, R.H.G., Lima, W.S., Cunha, M.C., 1982. *Schistosoma mansoni*: experimental bovine Schistosomiasis. Rev. Inst. Med. Trop. 24, 374–377.
- Coelho, P.M.Z., Lima, W.S., Nogueira, R.H.G., 1989. *Schistosoma mansoni*: on the possibility of Indian buffalo (*Bubalus bubalis*) being experimentally infected. Rev. Soc. Bras. Med. Trop. 22, 157–158.
- Coelho, P.M.Z., Lima, W.S., Modena, C.M., 1990. Activity of oxamniquine (Mansil) in experimental Schistosomiasis mansoni of calves. Arq. Bras. Med. Vet. Zootec. 42, 7–12.

- Combes, C., Fournier, A., Moné, H., Théron, A., 1994. Behaviours in trematode cercariae that enhance parasite transmission: patterns and process. *Parasitology* 30, 299–304.
- D'Andrea, P.S., Gentile, R., Cerqueira, R., Grelle, C.E.V., Horta, C., Rey, L., 1999. Ecology of small mammals in a Brazilian rural area. *Rev. Bras. Zool.* 16, 611–620.
- D'Andrea, P.S., Gentile, R., Maroja, L.S., Maldonado Jr., A., Cerqueira, R., Rey, L., 2000. The parasitism of *Schistosoma mansoni* (Digenea-Trematoda) in a naturally infected population of water-rat *Nectomys squamipes* (Rodentia-Sigmodontinae) in Brazil. *Parasitology* 120, 573–582.
- D'Andrea, P.S., Fernandes, F.A., Cerqueira, R., Rey, L., 2002. Experimental evidence and ecological perspectives for the adaptation of *Schistosoma mansoni* Sambon, 1907 (Digenea: Shistosomatidae) to a wild host, the water-rat, *Nectomys squamipes* Brants 1827 (Rodentia: Sigmodontinae). *Mem. Inst. Oswaldo Cruz.* 97, 11–14.
- Fenwick, A., 1969. Baboons as reservoir hosts of *Schistosoma mansoni*. *Trans. R. Soc. Trop. Med. Hyg.* 63, 557–567.
- Gentile, R., D'Andrea, P.S., Cerqueira, R., Maroja, L.S., 2000. Population and reproduction of marsupials and rodents in a Brazilian rural area: a five-year study. *Stud. Neotrop. Fauna & Environ.* 35, 1–9.
- Karoum, K.O., Amin, M.A., 1985. Domestic and wild animals naturally infected with *Schistosoma mansoni* in the Gezira irrigate scheme. *Sudan. J. Trop. Med. Hyg.* 88, 83–90.
- Keeling, M.J., Wilson, H.B., Pacala, S.W., 2000. Reinterpreting space, time lags, and functional responses in ecological models. *Science* 290, 1758–1761.
- Kloos, H., Gazzinelli, A., Van Zuylen, P., 1998. Microgeographical patterns of schistosomiasis and water contact behavior; examples from Africa and Brazil. *Mem. Inst. Oswaldo Cruz.* 93, 37–50.
- Koskella, B., Lively, C.M., 2007. Advice of the rose: experimental coevolution of a trematode parasite and its snail host. *Evol. Int. J. Org. Evol.* 61, 152–159.
- Kristt, A.C., Lively, C.M., Levri, E.P., Jokela, J., 2000. Spatial variation in susceptibility to infection in a snail-trematode interaction. *Parasitology* 121, 395–401.
- Mahmoud, A.A., 2004. Schistosomiasis (bilharziasis): from antiquity to the present. *Infect. Dis. Clin. North. Am.* 18, 207–218.
- Martins, A.V., Martins, G., Brito, R.S., 1955. Reservatórios silvestres do *Schistosoma mansoni* no Estado de Minas Gerais. *Rev. Bras. Malariol. Doenças Trop.* 7, 259–265.
- Mayaudon, H.T., Power, L.A., 1970. Infestacion natural de bovinos (*Bos taurus*) de Venezuela por *Schistosoma mansoni*. *Rev. Med. Parasitol.* 23, 1–8.
- McCully, R.M., Kuger, S.P., 1969. Observations on Bilharziasis on domestic ruminants in South Africa. *Onderstepoort J. Vet. Res.* 36, 129–162.
- Millward de Andrade, R., Carvalho, O.S., Cortes, M.I.N., 1976. Ciclo vital do *Schistosoma mansoni* através do *Holochilus brasiliensis* (Desmaret), em ambiente semi-natural. *Rev. Bras. Med. Trop.* 10, 235–247.
- Modena, C.M., Lima, W.S., Coelho, P.M.Z., Barbosa, F.S., 1991. Aspectos epidemiológicos da esquistossomose mansoni em bovinos. *Arch. Bras. Med. Vet. Zootec.* 43, 481–488.
- Modena, C.M., Coelho, P.M.Z., Barbosa, F.S., Lima, W.S., 1993. Transmission of *Schistosoma mansoni* under experimental conditions using the bovine-*Biomphalaria glabrata*-bovine model. *Rev. Inst. Med. Trop. São Paulo* 35, 11–16.
- Morgan, J.A., DeJong, R.J., Lwambo, N.J., Mungai, B.N., Mkoji, G.M., 2003. First report of a natural hybrid between *Schistosoma mansoni* and *S. rodhaini*. *J. Parasitol.* 89, 416–418.
- Nelson, G.S., 1960. *Schistosoma* infections as zoonoses in Africa. *Trans. R. Soc. Trop. Med. Hyg.* 54, 301–316.
- Ocampo, E.M., Coelho, P.M.Z., Lima, W.S., Nogueira, R.H.G., 1981. *Schistosoma mansoni* Infecção experimental em suínos (*Sus scrofa*). *Arq. Esc. Vet.* 33 (3), 467–470.
- Osnas, E.E., Lively, C.M., 2004. Parasite dose, prevalence of infection and local adaptation in a host-parasite system. *Parasitology* 128, 223–228.
- Ovaskainen, O., Cornell, S.J., 2006. Space and stochasticity in population dynamics. *Proc. Natl. Acad. Sci.* 103, 12781–12786.
- Pitchford, R.J., Visser, P.S., 1962. The role of naturally infected wild rodents in the epidemiology of schistosomiasis in the eastern Transvaal. *Trans. R. Soc. Trop. Med. Hyg.* 56, 126–135.
- Piva, N., Barros, P.R.C., 1966. Infecção natural de animais silvestres e domésticos pelo *Schistosoma mansoni* em Sergipe. *Rev. Bras. Malariol. Doenças Trop.* 18, 221–233.
- Poulin, R., 2007. Are there general laws in parasite ecology? *Parasitology* 19, 1–14.
- Saede, A.D., Nelson, G.S., Hussen, M.F., 1969. Experimental infections of calves with *Schistosoma mansoni*. *Trans. Roy. Soc. Trop. Med. Hyg.* 63, 456–458.
- Schwet, J., 1953. On a new schistosome of wild rodents found in Belgian Congo, *Schistosoma mansoni* var. *rodentorum*, var. nov. *Ann. Trop. Med. Parasitol.* 47, 183–186.
- Silva, R.R., Machado-Silva, J.R., Faerstein, N.F., Lenzi, H.L., Rey, L., 1992. Natural infection of wild rodents by *Schistosoma mansoni*: parasitological aspects. *Mem. Inst. Oswaldo Cruz.* 87, 271–276.
- Sire, C., Durand, P., Pointier, J.P., Théron, A., 2001. Genetic diversity of *Schistosoma mansoni* within and among individual hosts (*Rattus rattus*): infrapopulation differentiation at microspatial scale. *Inter. J. Parasitol.* 31, 1609–1616.
- Stothard, J.R., Lockyer, A.E., Kabatereine, N.B., Tukahebwa, E.M., Kazibwe, F., Rollinson, D., Fenwick, A., 2004. *Schistosoma bovis* in western Uganda. *J. Helminthol.* 78, 281–284.
- Théron, A., 1984. Early and late shedding patterns of *Schistosoma mansoni* cercariae: ecological significance in transmission to human and murine hosts. *J. Parasitol.* 70, 652–655.
- Théron, A., 1989. Hybrids between *Schistosoma mansoni* and *S. rodhaini*: characterization by cercarial emergence rhythmicity. *Parasitology* 99, 225–228.
- Théron, A., Pointier, J.P., 1995. Ecology, dynamics, genetics and divergence of trematode population in heterogeneous environments: the model of *Schistosoma mansoni* in the insular focus of Guadeloupe. *Res. Rev. Parasitol.* 55, 49–64.